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Feedback on XVI ASF Inter-laboratory Comparison Test (ILCT) 2019

Dear Dr. Loeffen:

This is to confirm the participation of the **National Veterinary Research Institute [NVRI], Poland, (laboratory designation code 23)** in the XVI ILCT 2018-2019 for African Swine fever disease (ASF), organised by the European Union Reference Laboratory (EURL) for ASF with the support of DG SANTÉ. The panel of samples included 14 serum samples, coded as S1 to S14, and 6 tissue samples, coded as T1 to T6, which were distributed for testing the presence of ASF.

The test results obtained by the different participating laboratories will be presented and discussed at the "**Workshop on Laboratory Diagnosis of African and Classical Swine Fever (ASF and CSF)"** that will be held in Madrid, Spain, at 17th,18th June 2019, with the participation of the EU´s National Swine Fever Reference Laboratories (NRLs) and third countries. A final report of this Meeting will be available at the EURL web page on July 2019.

A detailed report about the analyses of your results is attached in the annexed *23-ASF$_{ILCT19}$ report.* Comments and recommendations for each test that your laboratory performed for the ASF ILCT 2019 are showed below:

1. **ASF antibody detection results:** your laboratory used the commercial ELISA ®INGENASA PPA COMPAC 1.1 PPA k3 and the IDVET- indirect ELISA kit (ID Screen® African Swine Fever Indirect) for ASF antibody detection in serum samples. The indirect immunoperoxidase (IPT) and the Immunoblotting (IB) were used as confirmatory tests. **Your results were correct and 'as expected' in positive and negative serum samples indicating that the assay systems that you are using are 'fit for purpose' for the detection of antibodies against ASFV.**

2. **ASF virus detection results:** your laboratory used three real time PCR methods, i) the UPL-real time – PCR, ii) the commercial real time PCR "Virotype® ASFV PCR Kit Qiagen", and iii) the ID Gene™ African Swine Fever Duplex, IDVET GENETISC. Different extraction methods

were assayed comprising the QIAmp DNA Mini Kit and The High Pure PCR Template. **Your results were correct and 'as expected' in serum and tissue samples indicating that the assay systems that you are using are 'fit for purpose' for the detection of the ASF virus.** Different results obtained in weak positive serum samples have not been considered since a correct ASF final diagnostic conclusion has been provided combining both ASF virus and antibody detection tests.

The ASF final diagnostic conclusion provided in each of the samples included in the XVI ASF ILCT 2019 has been correct and in line with our expectations. From these results the EU Reference Laboratory for ASF informs that the diagnostic procedures that you are using are 'fit for purpose' to give a correct diagnosis of ASF.

Please contact us if you feel the results for your laboratory have been incorrectly interpreted. Furthermore, also contact us if you require any further information or assistance regarding recommended follow-up and corrective actions arising from the ILCT.

In Valdeolmos, Madrid, Spain, at 4th April 2019

Yours sincerely,

Dr. Carmina Gallardo,
Researcher, Laboratory Coordinator
EU reference laboratory for ASF
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